

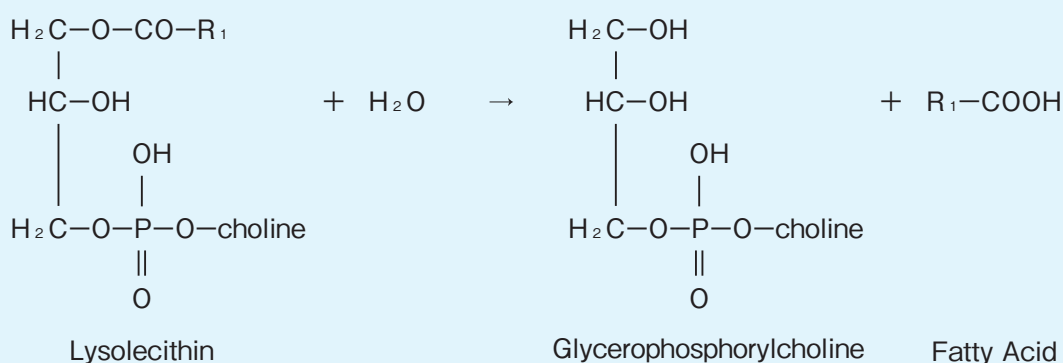
(Diagnostic Reagent Grade)

INQUIRY NEEDED
ASAHI KASEI ENZYMES T-32

LYSOPHOSPHOLIPASE [LYPL]

from *Vibrio* sp.

(2-Lysophosphatidylcholine acylhydrolase, EC 3.1.1.5)



Preparation and Specification

Appearance : White to brownish amorphous powder, lyophilized
 Specific activity : More than 20 U/mg solid

Properties

Michaelis constant	: Lysolecithin 6.7×10^{-4} M	
Optimum pH	: 9.0-9.5	Figure 1
pH stability	: 6.5-9.0 (37°C, 60 min)	Figure 2
Thermal stability	: Stable at 60°C and below (pH 7.2, 10 min)	Figure 3
Effect of various chemicals	: See Table 1 and Table 2	
Activator	: Ca^{2+}	
Inhibitors	: Cationic detergents	

Applications for Diagnostic Test

This enzyme is useful for enzymatic determination of **lysolecithin** when coupled with glycerophosphorylcholine phosphodiesterase (T-33) and choline oxidase (T-05).

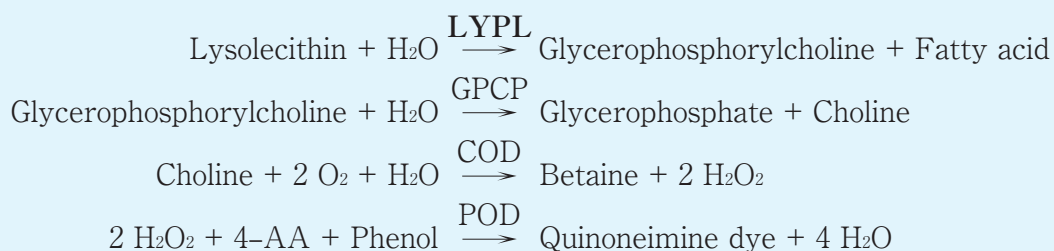
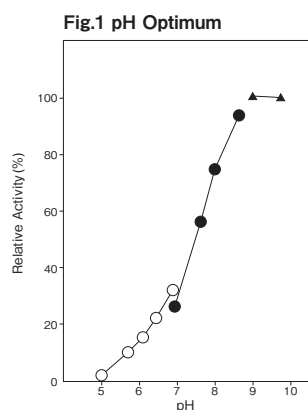


Table 1. Effect of detergents on lysophospholipase activity

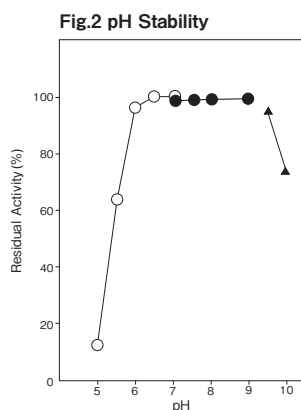
Detergent	Concentration (%)	Relative activity (%)
None	-	100
Triton X-100	0.5	119
Adekatorl SO-145	0.5	42
Adekatorl PC-8	0.5	74
Adekatorl NP-700	0.5	103
Pluronic L-61	0.5	119
Sodium cholate	0.5	117
Cethyl pyridinium chloride	0.5	0
Cethyl trimethyl-Ammonium chloride	0.5	0

Table 2. Effect of metal ions on lysophospholipase activity

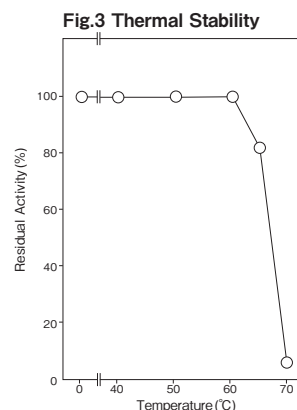
Metal ion	Concentration (mM)	Relative activity (%)
None	-	100
CaCl ₂	1	119
MgCl ₂	1	115
MnCl ₂	1	130
ZnCl ₂	1	14
BaCl ₂	1	127
CoCl ₂	1	117
CuCl ₂	1	9
NiCl ₂	1	110
NH ₄ Cl	100	71
NaCl	100	89
KCl	100	81



○ : 3,3-Dimethylglutarate
-NaOH buffer
● : Tris-HCl buffer
▲ : Borate buffer



37°C, 60 min.
○ : 3,3-Dimethylglutarate
-NaOH buffer
● : Tris-HCl buffer
▲ : Borate buffer

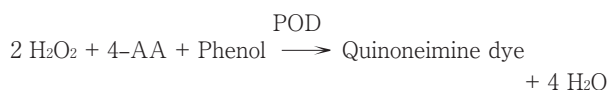
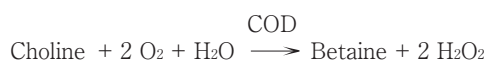
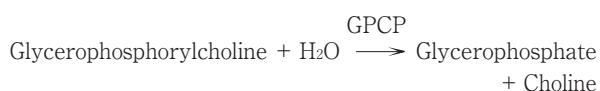


pH 7.2, 10 min.
10 mM Tris-HCl buffer

Assay

Principle

The assay is based on the increase in absorbance at 500 nm as the formation of quinoneimine dye proceeds in the following reactions:



GPCP: Glycerophosphorylcholine phosphodiesterase

COD: Choline oxidase

Unit definition

One unit is defined as the amount of enzyme which hydrolyzes 1 μmole of lysolecithin per minute at 37°C under the conditions specified in the assay procedure.

Reagents

1. Reaction mixture for the first reaction

0.2 M Tris-HCl buffer pH 8.0	0.10 ml
10 mM Lysolecithin solution	0.05 ml
10 mM CaCl ₂ solution	0.05 ml
1 U/ml GPCP solution ¹⁾	0.10 ml
Distilled water	0.15 ml

1): 1 U/ml GPCP solution

Dissolve 10 U of GPCP with 10 ml of 10 mM Tris-HCl buffer pH 8.0.

2. Reaction mixture for the second reaction

0.2 M Tris-HCl buffer pH 8.0	0.10 ml
0.1 M EDTA solution pH 8.0	0.20 ml
15 mM 4-AA solution	0.10 ml
0.2 % (W/V) Phenol solution	0.10 ml
60 U/ml COD solution ²⁾	0.10 ml
100 U/ml POD solution ³⁾	0.05 ml
Distilled water	0.35 ml

EDTA: Ethylenediamine tetraacetic acid

2): 60 U/ml COD solution

Dissolve 600 U of COD with 10 ml of 10 mM Tris-HCl buffer pH 8.0.

3): 100 U/ml POD solution

1,000 U (PPU) of POD with 10 ml of distilled water.

3. Enzyme dilution buffer
10 mM Tris-HCl buffer pH 8.0 containing 0.05% (W/V) bovine serum albumin

4. Reagents:

Lysolecithin (1-Oleoyl-sn-glycero-3-phosphocholine) :
Sigma Chemical Co. #L-1881
GPCP: Asahi Kasei Pharma Corporation #T-33
COD: Asahi Kasei Pharma Corporation #T-05
4-AA: NACALAI TESQUE, INC. Special grade #01907-52
POD: Sigma Chemical Co. Type II #P-8250
EDTA (2Na·2H₂O): KISHIDA CHEMICAL Co., Ltd.
#060-29133

■ Enzyme solution

Weigh about 20 mg of test sample exactly and add enzyme dilution buffer to make a total of 20 ml. Dilute it with enzyme dilution buffer to adjust the concentration as required.

■ Procedure

- Pipette 0.45 ml of reaction mixture for the first reaction into a small test tube and preincubate at 37°C.
- After 5 min, add exactly 50 μl of enzyme solution and mix to start the first reaction at 37°C.
※ In the case of a test blank, add 50 μl of enzyme dilution buffer in place of enzyme solution at this point.
- After 10 min, add immediately 1.0 ml of reaction mixture for the second reaction and mix to start the second reaction at 37°C.
- After 20 min, add 1.5 ml of distilled water to stop the reaction.

5. Measure the absorbance at 500 nm.

Absorbance sample : As
blank : Ab
 $\Delta A = (As - Ab) \leq 0.07$ Abs

■ Calculation

$$\text{Activity (U/mg)} = \frac{\Delta A/10}{12.0 \times 1/2} \times \frac{1}{2} \times \frac{3.00}{0.05} \times \frac{1}{X}$$

- 12.0 : millimolar extinction coefficient of quinoneimine dye at 500 nm (cm² / μmole)
2 : a multiplier derived from the fact that 1 mole of lysolecithin produces 2 mole of H₂O₂
1/2 : a multiplier derived from the fact that 2 mole of H₂O₂ produces 1 mole of quinoneimine dye
10 : reaction time (min)
3.00 : final volume (ml)
0.05 : volume of enzyme solution (ml)
X : concentration of the sample in enzyme solution (mg/ml)

Storage

Storage at -20°C in the presence of a desiccant is recommended.

References

- Misaki, H. and Matsumoto, M. (1978) J. Biochem., **83**, 1395-1405.
- Brumley, G. and Van Den Bosch, H. (1977) J. Lipid Res., **18**, 523-533.
- Scandella, C. J. and Kornberg, A. (1971) Biochemistry, **10**, 4447-4457.

LYPL 活性測定法 (Japanese)

I. 試薬液

1. 第一反応試薬混合液

0.2M トリス-HCl 緩衝液 pH8.0	0.10 ml
10mM リゾレシチン溶液	0.05 ml
10mM 塩化カルシウム溶液	0.05 ml
1U/ml GPCP 溶液 ¹⁾	0.10 ml
精製水	0.15 ml

1) : 1U/ml GPCP 溶液
GPCP 10 単位 (U) を 10mM トリス-HCl 緩衝液 pH8.0 10ml で溶解する。

2. 第二反応試薬混合液

0.2M トリス-HCl 緩衝液 pH8.0	0.10 ml
0.1M EDTA 溶液 pH8.0	0.20 ml
15mM 4-AA 溶液	0.10 ml
0.2% (W/V) フェノール液	0.10 ml
60U/ml COD 溶液 ²⁾	0.10 ml
100U/ml POD 溶液 ³⁾	0.05 ml
精製水	0.35 ml

2) : 60U/ml COD 溶液
COD 600 単位 (U) を 10mM トリス-HCl 緩衝

液 pH8.0 10ml で溶解する。

3) : 100U/ml POD 溶液

POD 1,000 単位 (PPU) を精製水 10ml で溶解する。

3. 酵素溶解希釈用液

0.05% (W/V) BSA を含む 10mM トリス-HCl 緩衝液 pH8.0

4. 試薬

リゾレシチン (1-オレオイル-sn-グリセロ-3-ホスホコリン): シグマ製 #L-1881

GPCP: 旭化成ファーマ製 #T-33

COD: 旭化成ファーマ製 #T-05

4-AA: ナカライテスク製 特級 #01907-52

POD: シグマ製 Type II #P-8250

EDTA (エチレンジアミン四酢酸·2Na·2H₂O): キシダ化学製 #060-29133

II. 酵素試料液

検品約 20mg を精密に量り、酵素溶解希釈用液で全容 20ml とする。

その液を酵素溶解希釈用液で適宜希釈する。

Ⅲ. 測定操作法

1. 小試験管に第一反応試薬混合液 0.45ml を正確に分注し、37℃で予備加温する。
2. 5分経過後、酵素試料液 50 μ l を正確に加えて混和し、37℃で第一反応を開始する。
※盲検は酵素試料液の代わりに酵素溶解希釈用液 50 μ l を加える。
3. 10分経過後、第二反応試薬混合液 1.0ml を加えて混和し、37℃で第二反応を開始する。
4. 20分経過後、精製水 1.5ml を加えて混和し、反応を停止する。
5. 500nm における吸光度を測定する。
求められた吸光度を試料液は A_s 、盲検液は A_b とする。

$$\Delta A = (A_s - A_b) \leq 0.07 \text{ Abs}$$

Ⅳ. 計算

$$\text{活性 (U/mg)} = \frac{\Delta A / 10}{12.0 \times 1/2} \times \frac{1}{2} \times \frac{3.00}{0.05} \times \frac{1}{X}$$

12.0 : キノンイミン色素の 500nm におけるミリモル分子吸光係数 ($\text{cm}^2 / \mu\text{mole}$)

1/2 : H_2O_2 2 モルからキノンイミン色素 1 モルが生成することによる係数

2 : リゾレシチン 1 モルから H_2O_2 2 モルが生成することによる係数

10 : 反応時間 (min)

3.00 : 反応総液量 (ml)

0.05 : 反応に供した酵素試料液量 (ml)

X : 酵素試料液の検品濃度 (mg/ml)