

SARCOSINE OXIDASE [SOXG]

from *Bacillus* sp.
(Sarcosine: oxygen oxidoreductase, EC 1.5.3.1)



Preparation and Specification

Appearance : Yellowish amorphous powder, lyophilized
Specific activity : More than 30 U/mg solid

Properties

Substrate specificity	: See Table 1	
Molecular weight	: 41 kDa (gel filtration)	
Isoelectric point	: pH 4.8	
Michaelis constant	: Sarcosine 3.4×10^{-2} M	
Optimum pH	: 7.5–8.5	Figure 1
pH stability	: 8.0–9.5 (50°C, 10 min)	Figure 2
Optimum temperature	: 45–50°C (pH8.0, 20mM Tris-HCl buffer)	Figure 3
Thermal stability	: Stable at 45°C and below (pH 7.5, 10 min)	Figure 4
Effect of various chemicals	: See Table2, Table3	

Applications for Diagnostic Test

This enzyme is useful for enzymatic determination of **creatinine** when coupled with creatinase and creatininase.

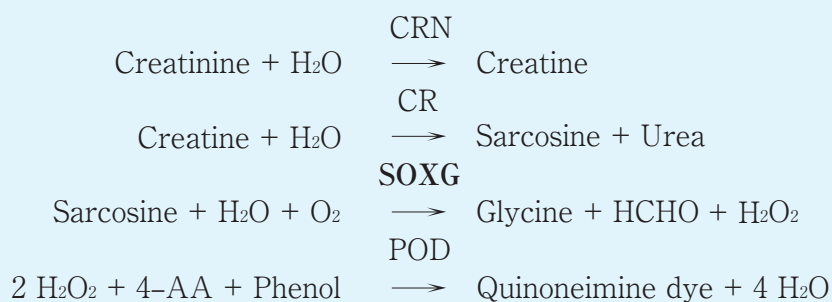


Table 1. Substrate specificity

Substrate	Relative activity (%)
Sarcosine	100
N-ethylglycine	11
Formylglycine	0
N,N-dimethylglycine	0
Glycine	0
Proline	0

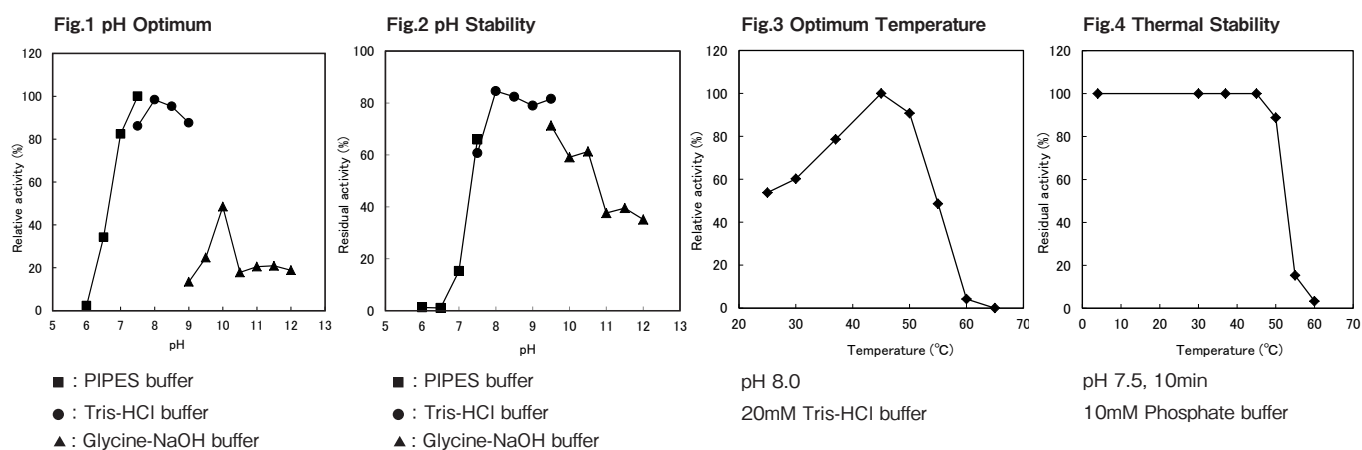
Table 2. Effect of various chemicals on SOXG stability (55°C, 10 min)

Additive	Concentration	Residual activity (%)
None		41
FAD	20 μM	30
KCl	0.3M	101
FMN	10 μM	21
EDTA	1mM	20
Sucrose	20%	80
Ethylene glycol	20%	2
Glycerol	20%	61

FAD: Flavin adenine dinucleotide
FMN: Flavin mononucleotide

Table 3. Effect of various chemicals on SOXG activity

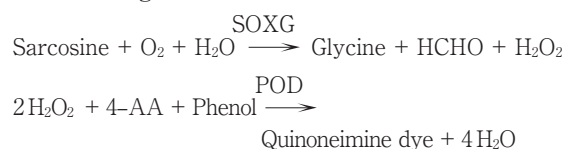
Additive	Concentration	Relative activity (%)
None		100
MgCl ₂	0.5mM	99
MnCl ₂	0.5mM	102
CaCl ₂	0.5mM	97
LiCl ₂	0.5mM	96
CuCl ₂	0.5mM	94
Ba(CH ₃ COO) ₂	0.5mM	100
NaCl	0.5mM	98
CoCl ₂	0.5mM	76
FeCl ₂	0.5mM	76
KCl	0.5mM	96
EDTA	1.0mM	98
Triton X-100	0.1%	99
Sodium Cholate	0.1%	92
Tween 80	0.1%	102



Assay

Principle

The assay is based on the increase in absorbance at 480 nm as the formation of quinoneimine dye proceeds in the following reactions:



Unit definition

One unit is defined as the amount of enzyme which oxidizes 1 μmole of sarcosine to glycine per minute at 37°C under the conditions specified in the assay procedure.

Reagents

- Reaction mixture

0.2M Tris-HCl buffer pH 8.0	0.05 ml
1.0M Substrate solution (Sarcosine)	0.10 ml
100U/ml POD solution ¹⁾	0.025 ml
15mM 4-AA solution	0.05 ml
0.2% (W/V) Phenol solution	0.05 ml
Distilled water	0.225 ml

1): 100 U/ml POD solution
Dissolve 1,000 U (PPU) of POD with 10 ml of distilled water.
- Reaction stopper
Ethanol
- Enzyme dilution buffer
10 mM KH₂PO₄-K₂HPO₄ buffer pH 7.5
- Reagents
Sarcosine (N-methylglycine or methylaminoacetate):
Tokyo Kasei Kogyo Co., Ltd. Special grade #M0332

4-AA: NACALAI TESQUE, INC. Special grade #01907-52
POD: Sigma Chemical Co. Type II # P-8250

■ Enzyme solution

Accurately weigh about 20 mg of the sample and add enzyme dilution buffer to make a total of 20 ml. Dilute it with enzyme solution buffer to adjust the concentration as required.

■ Procedure

1. Pipette accurately 0.5 ml of reaction mixture into a small test tube and preincubate at 37°C.
2. After 5 min, add exactly 10 μ l of enzyme solution and mix to start the reaction at 37°C.
※ In the case of a test blank, add 10 μ l of enzyme dilution buffer in place of enzyme solution.
3. At 5 min after starting the reaction, add 2.50 ml of the reaction stopper to stop the reaction.
4. Measure the absorbance at 480 nm.

$$\begin{aligned} \text{Absorbance sample} &: A_s \\ \text{Absorbance blank} &: A_b \\ \Delta A &= (A_s - A_b) \leq 0.125 \text{ Abs} \end{aligned}$$

■ Calculation

$$\text{Activity (U/mg of powder)} = \frac{\Delta A/5}{17.14 \times 1/2} \times \frac{3.01}{0.01} \times \frac{1}{X}$$

17.14 : millimolar extinction coefficient of quinoneimine dye at 480 nm ($\text{cm}^2 / \mu\text{mole}$)

1/2 : a multiplier derived from the fact that 2 mole of H_2O_2 produces 1 mole of quinoneimine dye

5 : reaction time (min)

3.01 : final volume (ml)

0.01 : volume of enzyme solution (ml)

X : concentration of the sample in enzyme solution (mg/ml)

Storage

Storage at -20°C in the presence of a desiccant is recommended. The enzyme activity will be retained for at least one year under this condition.

References

1. Mori, N., Sano, M., Tani, Y. and Yamada, H. (1980) *Agric. Biol. Chem.*, **44**, 1391-1397.
2. Suzuki, M. and Yoshida, M. (1976) *Proceedings of the Symposium on Chemical Physiology and Pathology (Kyoto)*, **Vol. 16**, 220.
3. Suzuki, M. (1981) *J. Biochem.*, **89**, 599-607.
4. Kinoshita, T. and Hiraga, Y. (1980) *Chem. Pharm. Bull.*, **28**, 3501-3506.

SOXG 活性測定法 (Japanese)

I. 試薬液

1. 反応試薬混合液
0.2M トリス-HCl 緩衝液 pH8.0 0.05 ml
1M 基質溶液 (サルコシン) 0.10 ml
15mM 4-AA 溶液 0.05 ml
0.2% (W/V) フェノール液 0.05 ml
100U/ml POD 溶液¹⁾ 0.025 ml
精製水 0.225 ml

1): 100U/ml POD 溶液

POD 1,000 単位 (PPU) を精製水 10ml で溶解する。

2. 反応停止液
エタノール原液を用いる。

3. 酵素溶解希釈用液
10mM KH_2PO_4 - K_2HPO_4 緩衝液 pH7.5

4. 試薬
サルコシン (N-メチルグリシン又はメチルアミノ酢酸): 東京化成製 特級 #M0332
4-AA: ナカライテスク製 特級 #01907-52
POD: シグマ製 Type II #P-8250

II. 酵素試料液

検品約 20mg を精密に量り、酵素溶解希釈用液で全容 20ml とする。

その液を酵素溶解希釈用液で適宜希釈する。

III. 測定操作法

1. 小試験管反応試薬混合液 0.50ml を正確に分注し、 37°C で予備加温する。
2. 5 分経過後、酵素試料液 10 μ l を正確に加えて混和し、 37°C で反応を開始する。

※ 盲検は酵素試料液の代わりに酵素溶解希釈用液 10 μ l を加える。

3. 5 分経過後、反応停止液 2.50ml を加えて混和し、反応を停止する。

4. 480nm における吸光度を測定する。

求められた吸光度の試料液は A_s 、盲検液は A_b とする。

$$\Delta A = (A_s - A_b) \leq 0.125 \text{ Abs}$$

IV. 計算

$$\text{活性 (U/mg)} = \frac{\Delta A/5}{17.14 \times 1/2} \times \frac{3.01}{0.01} \times \frac{1}{X}$$

17.14: キノンイミン色素の 480nm におけるミリモル分子吸光係数 ($\text{cm}^2 / \mu\text{mole}$)

1/2 : H_2O_2 モルからキノンイミン色素 1 モルが生成することによる係数

5 : 反応時間 (min)

3.01 : 反応総液量 (ml)

0.01 : 反応に供した酵素試料液量 (ml)

X : 酵素試料液の検品濃度 (mg/ml)