

# ASCORBATE OXIDASE [ASOM]

from *Acremonium* sp.  
(L-Ascorbate: oxygen oxidoreductase, EC 1.10.3.3)



## Preparation and Specification

Appearance : Light blue amorphous powder, lyophilized  
Specific activity : More than 200 U/mg solid

## Properties

Substrate specificity	: See Table 1	
Molecular weight	: 80 kDa (gel filtration)	
Isoelectric point	: pH 4.0	
Michaelis constants	: Ascorbic acid (pH 7.0) $1.0 \times 10^{-4}$ M Ascorbic acid (pH 4.0) $3.8 \times 10^{-4}$ M	
Optimum pH	: 4.0–4.5 (Acetate buffer)	Figure 1
pH stability	: 6.0–9.0 (30°C, 24 hr)	Figure 2
Thermal stability	: Stable at 50°C and below (pH 7.0, 10 min)	Figure 3
Liquid stability	: See Figure 4	
Effect of metal ions	: See Table 2	
Stabilizers	: BSA, Mannitol	

## Applications for Diagnostic Test

This enzyme is useful for **avoidance from interference of ascorbic acid** on diagnostic assay such as blood, uric acid, TG, TC and creatinine.

Table 1. Substrate specificity

Substrate	Relative activity (%)
L-Ascorbate	100
l-Naphthol	0
Hydroquinone	0
Catechol	0
Pyrogallol	0

Table 2. Effect of metal ions on ASOM activity

Metal ion	Concentration	Relative activity (%)
None		100
KCl	10 mM	100
NaCl	10 mM	100
EDTA	1 mM	100

Fig.1 pH Optimum

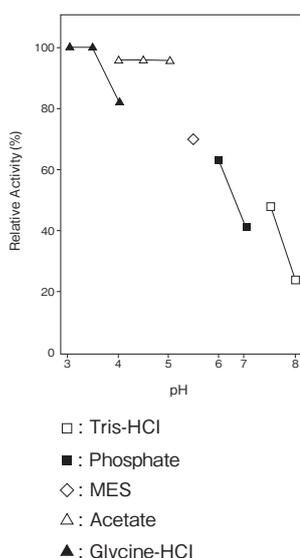


Fig.2 pH Stability

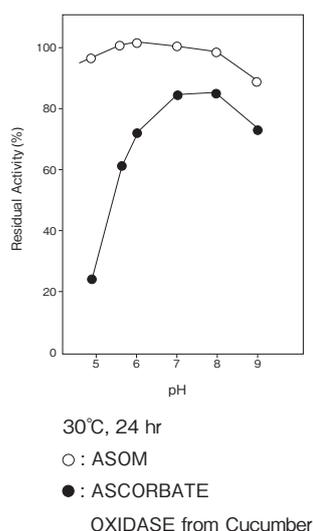


Fig.3 Thermal Stability

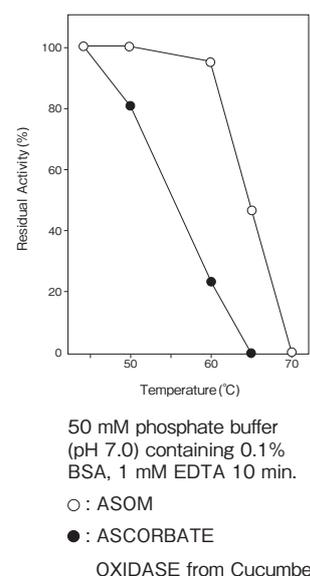
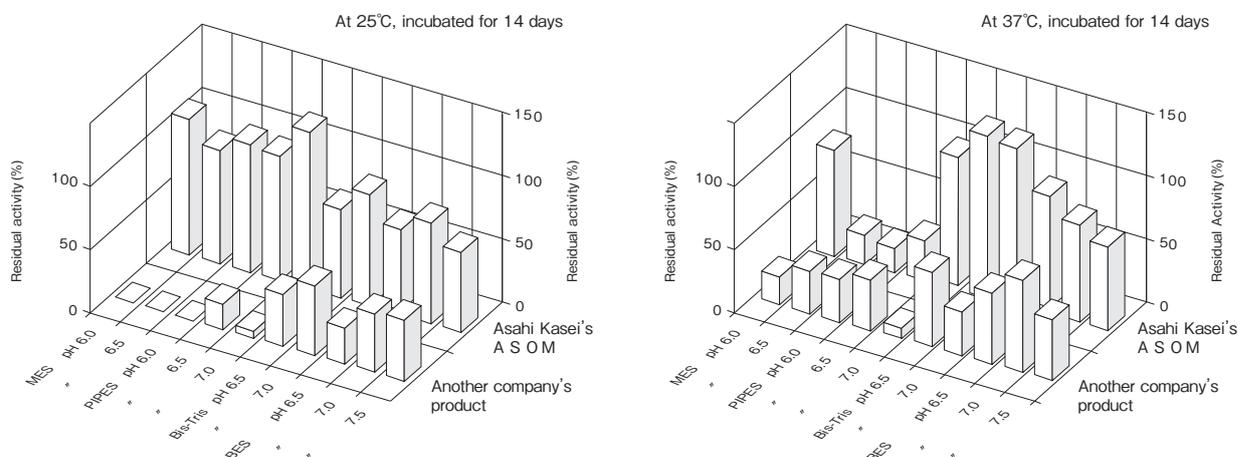


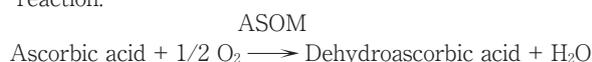
Fig.4 Liquid stability of ASOM (Buffer, pH)



## Assay

### Principle

The assay is based on the decrease in absorbance at 245 nm as ascorbic acid is oxidized in the following reaction:



### Unit definition

One unit is defined as the amount of enzyme which oxidizes 1 μmole of ascorbate to dehydroascorbate per minute at 30 °C under the conditions specified in the assay procedure.

## Reagents

### 1. Reaction mixture

Dilute substrate solution for stock<sup>1)</sup> with dilution buffer<sup>2)</sup> to make a 20-fold solution.

1): Substrate solution for stock (10 mM L-ascorbic acid solution)

Dissolve 176 mg of L-ascorbic acid and 37 mg of EDTA with 100 ml of 1mM HCl.

EDTA: Ethylenediamine tetraacetic acid

2): Dilution buffer

90mM KH<sub>2</sub>PO<sub>4</sub>-5mM Na<sub>2</sub>HPO<sub>4</sub> buffer containing 0.45 mM EDTA

2. Reaction stopper  
0.2 N HCl solution
3. Enzyme dilution buffer  
10 mM Na<sub>2</sub>HPO<sub>4</sub> solution containing 0.05% (W/V) BSA
4. Reagents

L-Ascorbic acid:

FUJIFILM Wako Pure Chemical Corporation  
Special grade # 012-04802

EDTA (2Na·2H<sub>2</sub>O): KISHIDA CHEMICAL Co., Ltd.  
#060-29133

BSA: Millipore Fraction V pH 5.2 #81-053

#### ■ Enzyme solution

Accurately weigh about 20 mg of the sample and add enzyme dilution buffer to make a total of 20 ml. Dilute it with enzyme dilution buffer to adjust the concentration to within 0.08-0.35 U/ml.

#### ■ Procedure

1. Pipette accurately 1.0 ml of reaction mixture into a small test tube and preincubate at 30°C.
2. After 5 min, add exactly 100 μl of enzyme solution and mix to start the reaction at 30°C.
3. At 5 min after starting the reaction, add 3.0 ml of the reaction stopper to stop the reaction.  
※ In the case of a test blank, add 100 μl of enzyme dilution buffer after adding reaction stopper in place of enzyme solution.

4. Measure the absorbance at 245 nm.

Absorbance sample : A<sub>s</sub>  
blank : A<sub>b</sub>

$$0.100A_{bs} \leq \Delta A = A_b - A_s \leq 0.420A_{bs}$$

#### ■ Calculation

$$\text{Activity (U/mg of powder)} = \frac{\Delta A/5}{10.0} \times \frac{4.10}{0.10} \times \frac{1}{X}$$

10.0 : millimolar extinction coefficient of ascorbic acid at 245 nm at pH 1.0 (cm<sup>2</sup>/μmole)

5 : reaction time (min)

4.10 : final volume (ml)

0.10 : volume of enzyme solution (ml)

X : concentration of the sample in enzyme solution

(mg/ml)

#### Storage

Storage at -20°C in the presence of a desiccant is recommended.

#### References

1. Murao, S., et al. (1991) Agric. Biol. Chem., **55** (6), 1693-1694.
2. Nakamura, T., Makino, N. and Ogura, Y. (1968) J. Biochem., **64**, 189.
3. Aikazyan, V. Ts. and Nalbandyan, R. M. (1979) FEBS Lett, **104**, 127.
4. White, G. A. and Smith, F. G. (1961) Nature, **190**, 187.

## ASOM 活性測定法 (Japanese)

### I. 試薬液

1. 保存基質溶液 (10mM L-アスコルビン酸)  
L-アスコルビン酸 176mg と EDTA 37mg を 1mM HCl 100ml で溶解する。
2. 反応試薬混合液  
上記の保存基質溶液を希釈用液<sup>※</sup>で 20 倍に希釈する。  
※): 希釈用液  
0.45mM EDTA を含む 90mM KH<sub>2</sub>PO<sub>4</sub>-5mM Na<sub>2</sub>HPO<sub>4</sub> 溶液
3. 反応停止液  
0.2N HCl 液
4. 酵素溶解希釈用液  
0.05% (W/V) BSA を含む 10mM Na<sub>2</sub>HPO<sub>4</sub> 溶液
5. 試薬  
L-アスコルビン酸:  
富士フィルム和光純薬製 特級 #012-04802  
EDTA (エチレンジアミン四酢酸·2Na·2H<sub>2</sub>O):  
キシダ化学製 #060-29133  
BSA: Millipore 製 Fraction V pH5.2 #81-053

### II. 酵素試料液

検品約 20mg を精密に量り、酵素溶解希釈用液に溶解して全容 20ml とする。

その液を酵素溶解希釈用液で 0.08~0.35U/ml 濃度となるように適宜希釈する。

### III. 測定操作法

1. 小試験管に反応試薬混合液 1.0ml を正確に分注して 30°C で予備加温する。
2. 5 分経過後、酵素試料液 100 μl を加えて混和し、30°C で反応を開始する。
3. 5 分経過後、反応停止液 3.0ml を加えて混和し、反応を停止する。  
※盲検は反応停止後に酵素試料液 100 μl を加える。
4. 245nm における吸光度を測定する。  
求められた吸光度を試料液は A<sub>s</sub>、盲検液は A<sub>b</sub> とする。

$$0.100A_{bs} \leq \Delta A = A_b - A_s \leq 0.420A_{bs}$$

### IV. 計算

$$\text{活性 (U/mg)} = \frac{\Delta A/5}{10.0} \times \frac{4.10}{0.10} \times \frac{1}{X}$$

10.0 : pH 1 の条件でアスコルビン酸の 245nm におけるミリモル分子吸光係数 (cm<sup>2</sup>/μmole)

5 : 反応時間 (min)

4.10 : 反応総液量 (ml)

0.10 : 反応に供した酵素試料液量 (ml)

X : 酵素試料液中の検品濃度 (mg/ml)